

This article was downloaded by:

On: 24 January 2011

Access details: *Access Details: Free Access*

Publisher *Taylor & Francis*

Informa Ltd Registered in England and Wales Registered Number: 1072954 Registered office: Mortimer House, 37-41 Mortimer Street, London W1T 3JH, UK



Journal of Liquid Chromatography & Related Technologies

Publication details, including instructions for authors and subscription information:

<http://www.informaworld.com/smpp/title~content=t713597273>

IMPROVED IDENTIFICATION BY IN SITU UV SPECTRA IN PLANAR CHROMATOGRAPHY

Anna Pelander^a; Ilkka Ojanperä^a; Johanna Sistonen^a; Pekka Sunila^b

^a Department of Forensic Medicine, University of Helsinki, Helsinki, Finland ^b Sunicom Oy, Helsinki, Finland

Online publication date: 31 May 2001

To cite this Article Pelander, Anna , Ojanperä, Ilkka , Sistonen, Johanna and Sunila, Pekka(2001) 'IMPROVED IDENTIFICATION BY IN SITU UV SPECTRA IN PLANAR CHROMATOGRAPHY', *Journal of Liquid Chromatography & Related Technologies*, 24: 10, 1425 – 1434

To link to this Article: DOI: 10.1081/JLC-100103920

URL: <http://dx.doi.org/10.1081/JLC-100103920>

PLEASE SCROLL DOWN FOR ARTICLE

Full terms and conditions of use: <http://www.informaworld.com/terms-and-conditions-of-access.pdf>

This article may be used for research, teaching and private study purposes. Any substantial or systematic reproduction, re-distribution, re-selling, loan or sub-licensing, systematic supply or distribution in any form to anyone is expressly forbidden.

The publisher does not give any warranty express or implied or make any representation that the contents will be complete or accurate or up to date. The accuracy of any instructions, formulae and drug doses should be independently verified with primary sources. The publisher shall not be liable for any loss, actions, claims, proceedings, demand or costs or damages whatsoever or howsoever caused arising directly or indirectly in connection with or arising out of the use of this material.

IMPROVED IDENTIFICATION BY IN SITU UV SPECTRA IN PLANAR CHROMATOGRAPHY

Anna Pelander,¹ Ilkka Ojanperä,^{1*} Johanna Sistonen,¹
and Pekka Sunila²

¹Department of Forensic Medicine, University of Helsinki,
P. O. Box 40, Kytösuontie 11, FIN-00014 Helsinki, Finland

²Sunicom Oy, P.O. Box 189, FIN-00931 Helsinki, Finland

ABSTRACT

Novel software was introduced for library searching in planar chromatography, based on corrected R_f values (hR_f^c), *in situ* ultraviolet (UV) spectral correlation, and spectrum maximum site comparison. Analysis of autopsy urine samples by thin-layer chromatography (TLC) and overpressured layer chromatography (OPLC) revealed that the new method was superior to the standard method using hR_f^c and correlation only. The advantage was more pronounced in TLC, which possessed poorer separation power and generally lower quality spectra than OPLC. In 11 cases of 47 in TLC, the hit list position of the correct finding was improved by the new method.

This result indicates that the spectrum maximum site function gives a different kind of discrimination than would be obtained by only setting the correlation cut-off at a higher value. In OPLC, the hit lists were originally shorter, but an improvement in the list

*Corresponding author.

lengths was evident. OPLC proved to be the planar method of choice in the screening analysis of urine samples for drugs due to its high separation power and speed.

INTRODUCTION

Thin-layer chromatography (TLC) in the instrumental mode utilizing corrected R_f values (hR_f^c),¹ measurement of *in situ* ultraviolet (UV) spectra, and special software for identification with substance libraries, has been available for screening analysis for several years.²⁻⁴ However, due to the limited separation power of TLC and the fairly low information content of UV spectra, long hit lists of candidates are often seen in the analysis report. In overpressured layer chromatography (OPLC), better separation numbers can be obtained compared to TLC. Recently, two complementary OPLC systems have been described for broad scale toxicological screening analysis for drugs.⁵

In spite of improved separation power, the hit lists may still remain relatively long. Use of the spectral correlation only seems to be inadequate to resolve between fairly similar UV spectra. In this study, an improved version of the identification software is described: the program uses spectrum maximum sites as an additional criterion for identification. The effect of the maximum site criterion on the length of hit lists is shown in TLC and OPLC analysis of drugs in urine.

EXPERIMENTAL

Materials and Apparatus

TLC

The chromatographic plates were 10 cm (height) x 20 cm (breadth) TLC glass plates coated with 0.25 mm layers of silica gel 60 F₂₅₄ (Merck, Darmstadt, Germany).

OPLC

The chromatographic plates were 20 cm x 20 cm HPTLC aluminium sheets coated with 0.2 mm layers of silica gel 60 F₂₅₄ (Merck). The plate edges were factory-sealed for OPLC use (OPLC-NIT Engineering Company, Budapest, Hungary). Drug standards were obtained from various pharmaceutical companies and were of pharmaceutical purity.

The standard and correction standard solutions were prepared in methanol to obtain a concentration of 2 mg/mL and 1 mg/mL, respectively. Analytical grade solvents were used throughout the study. The urine samples were collected at autopsy.

The automatic TLC sampler was an ATS III (Camag, Muttenz, Switzerland). The OPLC instrument was a Personal OPLC Basic System 50 (OPLC-NIT Engineering Company). The scanning densitometer was a TLC Scanner 3 (Camag) operated with CATS 4.03 software.

Chromatography

TLC

The mobile phase composition was toluene – acetone – 94% ethanol – 25% ammonia (45:45:7:3), and the chamber was allowed to saturate with a piece of filter paper for 0.5 h before development. The correction standards were codeine ($hR_f^c = 16$), promazine ($hR_f^c = 36$), clomipramine ($hR_f^c = 49$), and cocaine ($hR_f^c = 66$).² The drug library consisted of 355 substances measured with 4 μg amounts of pure substances.⁶

OPLC

The mobile phase composition was trichloroethylene – methylethylketone – *n*-butanol – acetic acid – water (17:8:25:6:4), and the development was carried out without pre-saturation of the plate. The external pressure was 50 bar, the flow-rate was 450 $\mu\text{L}/\text{min}$, the volume of rapid delivery was 300 μL , and the mobile phase volume was 5500 μL . The correction standards were codeine ($hR_f^c = 16$), promazine ($hR_f^c = 38$), nortriptyline ($hR_f^c = 58$), moperone ($hR_f^c = 76$), and theophylline ($hR_f^c = 86$).⁵ The drug library consisted of 113 substances measured with 1 μg amounts of pure substances.

Urine Analysis

The sample preparation for basic drugs in 5 mL urine specimens involved an ion-pair extraction with dichloromethane containing 0.01 M bis(2-ethylhexyl)phosphoric acid at pH 7.5.⁶ Aliquots of the reconstituted extracts, 10 μL for TLC and 1 μL for OPLC, and the correction standard mixture on a separate track, were applied band-wise to the plates with the automatic TLC sampler. The developed plates were evaluated by densitometry at 220 nm, and the *in situ* UV

Table 1. Hit Lists Obtained in the TLC Analysis of Urine Samples Using a) the Novel Software with hR_r^c + Spectrum Correlation + Spectrum Maximum Site Function (Method A) and by b) hR_r^c + Spectrum Correlation Only (Method B)

Number of Case	Correct Finding	a) Hit List Position of Correct Finding/ Hit List Length	b) Hit List Position of Correct Finding/ Hit List Length
H1034	Thioridazine	1 st /2	2 nd /11
2480	Zopiclone	1 st /1	1 st /4
2491	Amitriptyline	1 st /2	1 st /6
2499	Zopiclone	1 st /1	1 st /4
2520	Dextropropoxyphene	2 nd /5	4 th /12
2540	Levomepromazine	2 nd /5	2 nd /21
2553	Citalopram	1 st /5	1 st /24
2599	Quinine/Quinidine	1 st /1	1 st /3
2601	Quinine/Quinidine	1 st /1	1 st /4
2634	Citalopram	1 st /4	1 st /17
2647	Citalopram	1 st /6	1 st /18
2662	Nordoxepin	1 st /3	1 st /24
2674	Thioridazine	1 st /2	1 st /13
2701	Levomepromazine	2 nd /3	2 nd /17
2716	Citalopram	1 st /1	4 th /28
2759	Levomepromazine	2 nd /5	2 nd /11
2761	Quinine/Quinidine	1 st /1	1 st /4
2771	Orphenadrine	4 th /11	5 th /17
2784	Zopiclone	1 st /3	1 st /4
2801	Promazine	1 st /6	2 nd /20
2816	Citalopram	2 nd /6	2 nd /24
2823	Promazine	1 st /7	3 rd /27
2869	Orphenadrine	3 rd /7	3 rd /18
2889	Atenolol	1 st /5	1 st /15
2906	Metoclopramide	1 st /3	1 st /7
2931	Citalopram	1 st /5	1 st /18
2936	Promazine	2 nd /8	2 nd /22
2948	Amitriptyline	1 st /1	1 st /30
2948	Nortriptyline	1 st /1	1 st /19
2965	Metoprolol	4 th /10	4 th /20
3026	Sotalol	1 st /6	1 st /16
3054	Promazine	1 st /7	2 nd /20
3063	Zopiclone	1 st /1	2 nd /4
3085	Amphetamine	2 nd /6	2 nd /12
3098	Nordoxepin	1 st /2	1 st /19
3101	Doxepin	1 st /1	1 st /30
3109	Quinine/Quinidine	1 st /1	1 st /2

Table 1. Continued

Number of Case	Correct Finding	a) Hit List Position of Correct Finding/ Hit List Length	b) Hit List Position of Correct Finding/ Hit List Length
3132	Metoclopramide	1 st /1	2 nd /3
3134	Metformin	1 st /1	1 st /12
3134	Metoprolol	5 th /11	5 th /18
3193	Metoprolol	2 nd /7	2 nd /12
3218	Citalopram	1 st /5	1 st /13
3239	Ranitidine	1 st /1	1 st /5
3242	Zopiclone	1 st /2	1 st /4
3250	Promazine	1 st /3	3 rd /14
3252	Metoprolol	4 th /5	5 th /6
3272	Promazine	1 st /6	1 st /20

spectra were measured using a spectral range of 190-400 nm with 5 nm wavelength increments. CATS Spectrum Library for MS Windows (Camag) and a modified version of the software, developed in the present study, was used for the correction of R_f values and for the library search based on hR_f^c values and UV spectra.

RESULTS AND DISCUSSION

The novel software developed for substance identification in planar chromatography is an extension of the CATS Spectrum Library for MS Windows (Camag). In addition to hR_f^c and spectral correlation with substance libraries, identification within this program can also be based on the absorption maximum sites of the spectrum. It is also possible to be limited within a certain spectral region. In order to avoid too strict identification criteria, the number of maximum sites (Nmax) can be individually set at a lower number than the actual number for each library compound, and identification is based on any combination of maximum sites present in the sample spectrum.

The TLC library (355 substances) and OPLC library (133 substances) were updated by adding the spectrum maximum sites with wavelength windows to each substance individually. Based on preliminary experiments, Nmax was usually set at 1-3. For substances with one or two maximum sites in their spectrum, Nmax was given an equal value, but for substances with three or more maximum sites, Nmax was given a value of one less. The spectral region above 340 nm was

Table 2. Hit Lists Obtained in the OPLC Analysis of Urine Samples Using a) the Novel Software with hR_i^c + Spectrum Correlation + Spectrum Maximum Site Function (Method A) and by b) hR_i^c + Spectrum Correlation Only (Method B)

Number of Case	Correct Finding	a) Hit List Position of Correct Finding/ Hit List Length	b) Hit List Position of Correct Finding/ Hit List Length
2480	Zopiclone	1 st /1	1 st /1
2491	Amitriptyline	1 st /2	1 st /12
2491	Nortriptyline	1 st /1	1 st /6
2495	Diltiazem	1 st /6	1 st /10
2499	Zopiclone	1 st /1	1 st /1
2540	Chlorpromazine	1 st /2	1 st /6
2553	Citalopram	1 st /4	1 st /8
2599	Quinine/Quinidine	1 st /1	1 st /2
2634	Citalopram	1 st /2	1 st /3
2647	Citalopram	1 st /2	1 st /3
2662	Doxepin	1 st /1	1 st /7
2680	Citalopram	1 st /2	1 st /3
2680	Amitriptyline	1 st /3	1 st /12
2716	Zopiclone	1 st /1	1 st /1
2716	Citalopram	1 st /6	1 st /11
2716	Norcitalopram	1 st /2	1 st /9
2729	Chlorpromazine	1 st /1	1 st /2
2761	Quinine/Quinidine	1 st /1	1 st /1
2771	Orphenadrine	1 st /7	1 st /12
2784	Zopiclone	1 st /1	1 st /1
2816	Citalopram	1 st /5	1 st /8
2823	Promazine	1 st /2	1 st /3
2869	Nororphenadrine	1 st /7	1 st /17
2889	Atenolol	1 st /3	1 st /4
2906	Metoclopramide	1 st /1	1 st /2
2931	Citalopram	1 st /6	1 st /8
2931	Norcitalopram	1 st /2	1 st /8
2948	Amitriptyline	1 st /2	1 st /13
2948	Atenolol	1 st /3	1 st /4
2948	Nortriptyline	1 st /2	1 st /7
2950	Doxepin	1 st /1	1 st /3
3025	Sotalol	1 st /6	1 st /7
3032	Doxepin	1 st /1	1 st /6
3054	Promazine	1 st /1	1 st /5
3075	Doxepin	1 st /1	1 st /6
3085	Dextropropoxyphene	1 st /5	1 st /12
3098	Doxepin	1 st /1	1 st /10

Table 2. Continued

Number of Case	Correct Finding	a) Hit List Position of Correct Finding/ Hit List Length	b) Hit List Position of Correct Finding/ Hit List Length
3098	Nordoxepin	1 st /1	1 st /12
3101	Doxepin	1 st /1	1 st /9
3109	Quinine/Quinidine	1 st /1	1 st /1
3119	Atenolol	1 st /3	1 st /4
3132	Metoclopramide	1 st /1	1 st /2
3134	Metoprolol	1 st /11	1 st /14
3183	Diltiazem	1 st /3	1 st /6
3193	Metoprolol	1 st /12	1 st /14
3218	Citalopram	1 st /1	1 st /3
3239	Ranitidine	1 st /1	1 st /2
3239	Zopiclone	1 st /1	1 st /1
3239	Propranolol	1 st /1	1 st /1
3242	Zopiclone	1 st /1	1 st /1
3245	Amitriptyline	1 st /3	1 st /10
3247	Norcitalopram	1 st /1	1 st /2
3272	Promazine	1 st /2	1 st /5

rejected due to possible uncharacteristic maxima. Concentration differences and matrix effects of urine samples caused 1-3 nm shifts of the maximum sites, depending on the substance and type of spectrum, and thus a ± 3 nm wavelength window was set as a constant for all maximum sites of all library compounds.

Tables 1 and 2 compare the hit lists obtained by TLC and OPLC, respectively, using the spectrum maximum site function (Method A) and the standard software (Method B) in the analysis of urine samples. In these experiments, the hR_f^c window was ± 7 , and the spectrum correlation cut-off value was 0.8. The improvement was more pronounced in TLC, as could be expected on grounds of the lower separation power of the technique and the larger library size used. In 11 cases of 47 in TLC, the hit list position of the correct finding improved. This result indicates, that the spectrum maximum site function gives a different kind of discrimination than what would be obtained by only setting a higher correlation cut-off value. In OPLC, the hit lists were originally shorter, but an improvement in the list lengths was still evident. The chromatographic separation by TLC and OPLC in three selected cases is compared in Figures 1-3.

Although it has been possible to measure *in situ* UV spectra for more than thirty years,⁷⁻⁹ very little literature is available on the practical use of large UV spectrum libraries in planar chromatography. It has been stated that UV library

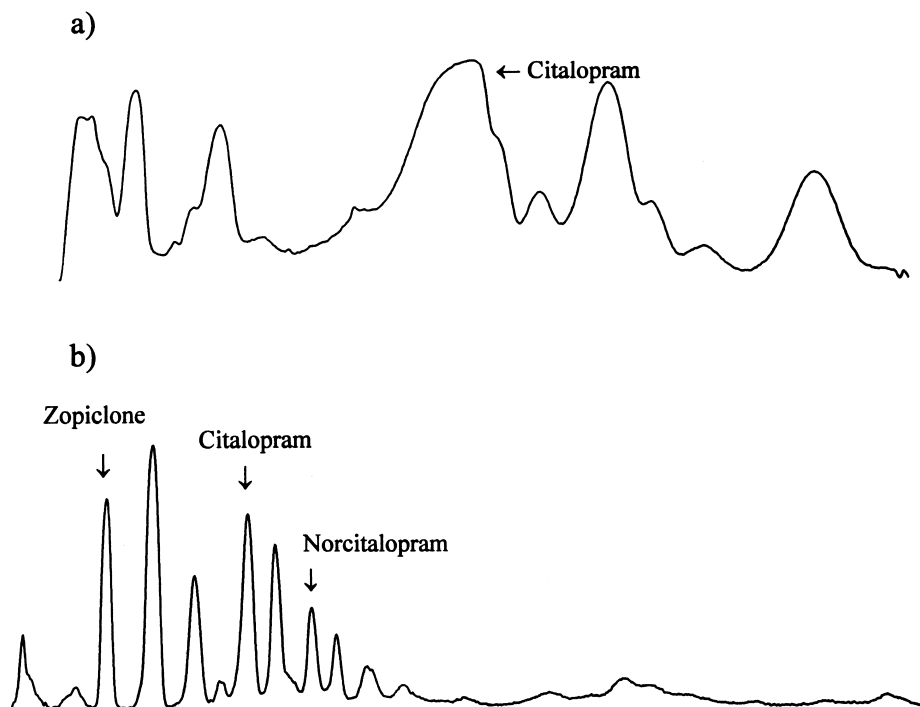


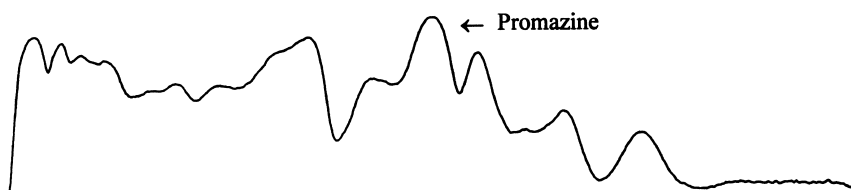
Figure 1. Chromatograms obtained from case 2716 urine sample by a) TLC with the finding citalopram, and b) OPLC with the findings zopiclone, citalopram and norcitalopram.

search procedures within planar techniques are not useful,¹⁰ but the recent work done in the authors' laboratory²⁻⁶ shows the opposite.

CONCLUSIONS

Novel software, which utilized both UV spectral correlation and spectrum maximum sites for identification, was found to be superior in terms of hit list length and validity of results, compared to the standard program using correlation only. This was especially pronounced in ordinary TLC, which exhibited poorer separation and generally lower quality spectra than OPLC. It is evident that OPLC possesses great potential in broad scale screening analysis for drugs due to

a)



b)

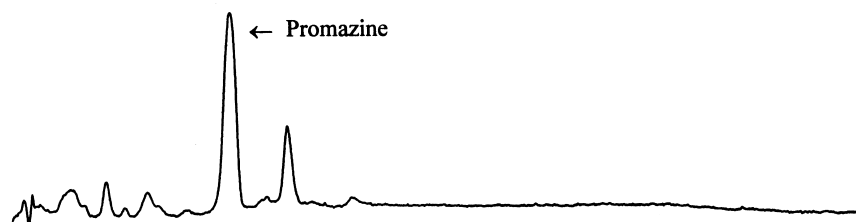
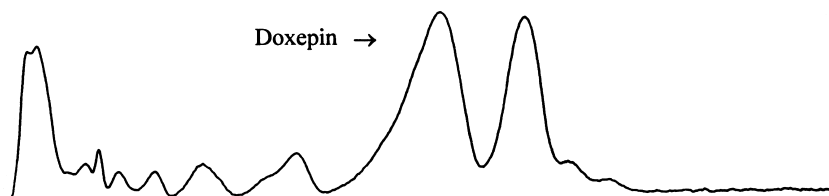


Figure 2. Chromatograms obtained from case 2823 urine sample by a) TLC and b) OPLC, with the finding promazine.

a)



b)

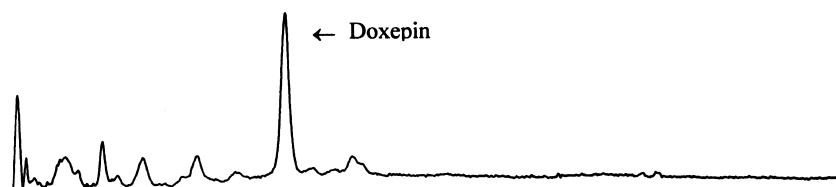


Figure 3. Chromatograms obtained from case 3101 urine sample by a) TLC and b) OPLC, with the finding doxepin.

its high separation power and speed, especially in connection with these new means of UV spectral discrimination.

REFERENCES

1. *Thin-Layer Chromatographic R_f Values of Toxicologically Relevant Substances in Standardized Systems*, 2nd Ed.; de Zeeuw, R.A., Franke, J.P., Degel, F., Schütz, H., Wijsbeek, J., Eds.; VCH: Weinheim, 1992.
2. Ojanperä, I.; Jänchen, P. *LC-GC Int.* **1994**, *7*, 164-170.
3. Ojanperä, I.; Vuori, E. *J. Chromatogr.* **1994**, *674*, 147-152.
4. Ojanperä, I.; Nokua, J.; Vuori, E.; Sunila, P., Sippola, E. *J. Planar Chromatogr.-Mod. TLC* **1997**, *10*, 281-285.
5. Ojanperä, I.; Goebel, K.; Vuori, E. *J. Liq. Chromatogr. Relat. Technol.* **1999**, *22*, 161-171.
6. Ojanperä, I. *Thin-Layer Chromatography in Forensic Toxicology*. In *Practical Thin-Layer Chromatography, A Multidisciplinary Approach*; Fried, B., Sherma, J., Eds.; CRC Press: Boca Raton, 1996; 193-230.
7. Ebel, S.; Kang, J.S. *J. Planar Chromatogr.-Mod. TLC* **1990**, *3*, 42-46.
8. Jork, H. *Am. Lab.* **June 1993**, 24B-24F.
9. Gocan, S.; Cimpan, G. *Rev. Anal. Chem.* **1997**, *16*, 1-24.
10. Poole, C.F.; Poole, S.K. *Anal. Chem.* **1994**, *66*, 27A-37A.

Received November 7, 2000
Accepted December 16, 2000

Manuscript 5451